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(54) Odorant-binding protein from rat.

(57) DNA molecules are taught which code for an odorant-binding protein which is synthesized solely in the lateral nasal gland. This protein, because of the broad range of odorants which it binds, can be used in many techniques for trapping odorants in either a liquid or solid medium. This protein bears some structural homology with other carriers of small lipophilic molecules from many other species; the carriers are known to transport specific lipophilic molecules.

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Description

ODORANT-BINDING PROTEIN FROM RAT

This work was partially supported by grants from the United States Department of Health and Human Services. The United States Government may retain certain rights in this invention.

TECHNICAL FIELD OF THE INVENTION

This invention relates to the area of olfactory perception. More specifically it relates to the initial steps in the process of olfactory perception.

BACKGROUND OF THE INVENTION

The complete molecular mechanism responsible for olfaction has yet to be elucidated. However, odorant binding activity has been detected in mammalian olfactory tissue extracts. In addition, behavioral and neurophysiological evidence suggests the existence of specific olfactory receptor proteins. For example, Amoore reports losses of the sense of smell (anosmias) to about thirty discrete odorant groups. (Chem. Senses Flavor, Vol. 2, pp. 267-281, 1977.) In addition, there is evidence for a binding protein for urinous odorants (Persaud et al, in *Biochemistry of Taste and Olfaction* (1981), Cagan et al, editors, pp. 333-357, Academic Press, New York), a camphor-binding protein (Fesenko et al, *Biochim. Biophys. Acta*, Vol. 587, pp. 424-433, 1979) and a binding protein for green smelling odorants such as 2-isobutyl-3-methoxyprazine (IBMP) (Pelosi et al, *Biochemical Journal*, Vol. 201, pp. 245-248, 1982). Despite these findings, the molecular identity of most of the components of the olfactory system remain undefined.

At the molecular level, several proteins have been identified which appear to be involved in olfaction. Lee et al, *Science*, Vol. 235, pp. 1053-1956 (1987) report the isolation of an olfactory cDNA from the frog. The mRNA corresponding to the isolated cDNA appears to be abundant in the Bowman's glands in olfactory tissue, but not in other tissues tested. Although the function of the protein encoded by the gene is unknown, its sequence shares homology with members of a family of proteins that bind and transport small molecules in the serum, suggesting a transporting role for this protein.

Another protein presumably involved in olfaction is the so-called pyrazine-binding protein or odorant-binding protein from cow nasal tissue. This protein was first detected on the basis of its binding activity for "green" smelling compounds such as 2-isobutyl-3-methoxyprazine. (See Bignetti et al, *European Journal of Biochemistry*, Vol. 149, pp. 227-231, 1985 and Pevsner et al, *Proceedings of the National Academy of Sciences*, Vol. 82, pp. 3050-3054, 1985.) This cow protein has been purified to apparent homogeneity and a partial amino acid sequence has been determined (See Cavaggioni et al, *FEBS Letters*, Vol. 212, pp. 225-228, 1987). Homology between this cow protein and the major mouse urinary proteins (MUP) and the rat alpha-2-microglobulins (AMG) was noted.

A rat odorant-binding protein has also been purified to apparent homogeneity. (Pevsner et al, *Proceedings of the National Academy of Sciences USA*, Vol. 83, pp. 4942-4946, 1986.) The protein was found in both olfactory and respiratory epithelium. Although it was known that the rat odorant protein was secreted into the nasal mucus and tears, it was not known where this protein was synthesized.

Another identified protein which is presumably involved in olfaction is the so-called olfactory marker protein (OMP). This is a neuronal gene product which is developmentally regulated and whose expression is restricted to mature olfactory receptor neurons of many vertebrate species. The gene for this protein has been cloned and sequenced. (See Rogers et al, *Proceeding of the National Academy of Sciences USA*, Vol. 84, pp. 1704-1708, 1987.) Another protein which has been identified in the olfactory perception system is an odorant-sensitive adenylate cyclase. This was found in olfactory sensory neurons and requires the presence of guanosine triphosphate (GTP). Only certain odorants are found to stimulate the enzyme activity. (See *Journal of Biological Chemistry*, Vol. 61, pp. 15538-15543, 1986.)

There is a need for a more thorough understanding of the components of the olfactory perception system. Molecular cloning of these components can facilitate the understanding of the mechanism of olfaction by allowing the production of large amounts of the individual components. Further, molecular cloning can facilitate the determination of the sites of synthesis of various components of the olfactory system.

Because odorant molecules are often present in rather small amounts in natural sources, they can be difficult to isolate and/or analyze. Thus there is a need for a large source of an odorant binder to facilitate the isolation and identification of odorant molecules. Further, some odorant molecules are readily absorbed into the skin which can both be deleterious to the health and cause the odorant (for example, a perfume) to have a short active life on the skin. Therefore there is a need in the art for compounds which can extend the active life of odorants on the skin. In addition, many odorants are lipophilic, and thus are difficult to bring into aqueous or semi-aqueous solution. Thus there is a need in the art for compounds which can facilitate the emulsification of lipophilic odorant molecules.

SUMMARY OF THE INVENTION

It is an object of the present invention to provide a method of isolating an odorant-binding protein gene.

It is another object of the invention to provide a method of producing an odorant-binding protein.

It is yet another object of the invention to provide a DNA molecule which encodes an odorant-binding

protein.

It is another object of the invention to provide a method of concentrating odorants on a solid support.
It is another object of the present invention to provide a method of reducing the rate of volatilization of odorants from solutions.

It is yet another object of the present invention to provide a method of solubilizing lipophilic odorants in aqueous media.

It is still another object of the present invention to provide a method of screening for potent odorants in crude extracts of natural tissues.

These and other objects are fulfilled by the present invention in one or more of the embodiments described below. In one embodiment of the present invention, a method is provided for isolating an odorant-binding protein gene, comprising: preparing cDNA using mRNA templates isolated from lateral nasal gland tissue; inserting the cDNA into DNA vectors to form recombinant DNA molecules; transforming host cells with said recombinant DNA molecules to form transformed cells; and detecting the presence of odorant-binding protein DNA in said transformed cells.

In another embodiment of the invention a method is provided for producing an odorant-binding protein, comprising: transforming host cells with a recombinant DNA molecule comprising an odorant-binding protein gene which is positioned downstream from a transcriptional promoter and downstream from and in phase with a translational start site to form transformed cells; growing said transformed cells in a culture medium under conditions such that said transformed cells express the protein encoded by said odorant-binding protein gene; and harvesting the expressed protein encoded by said odorant-binding protein gene from said transformed cells and/or said growth medium.

In yet another embodiment of the invention a DNA molecule is provided which is free of introns and which encodes a protein having a sequence corresponding to that shown in Figure 1 for the rat odorant-binding protein. The rat odorant-binding protein has been discovered to be synthesized in the lateral nasal glands (LNG). Such odorant-binding proteins which are produced in the LNG are termed herein OBP_L.

In yet another embodiment of the present invention a method is provided of concentrating odorants on a solid support, comprising: applying an OBP_L to a solid support, and contacting said solid support with odorant compounds to effect binding of the odorant to said solid support via OBP_L.

In yet another embodiment of the present invention a method is provided of reducing the rate of volatilization of odorants from solutions, comprising: providing a solution containing a volatile odorant, and mixing a volatilization-reducing amount of OBP_L with said solution.

In still another embodiment of the present invention a method is provided of solubilizing lipophilic odorants in aqueous media, comprising: providing an aqueous medium; binding a lipophilic odorant to OBP_L, and mixing said OBP_L-bound lipophilic odorant with the aqueous medium to form an aqueous perfumed product.

In yet another embodiment of the present invention a method is provided of screening for potent odorants in crude extracts of natural tissues, comprising: binding a strong odorant to OBP_L to form a bound OBP_L and strong odorant complex; making an extract of a natural tissue; making serial dilutions of the extract; and determining which dilutions of the extract are able to displace the bound, strong odorant from the complex, an extract which can displace the strong odorant at a high dilution indicating the presence of a potent odorant in the natural tissue.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the nucleotide sequence and predicted amino acid sequence of three cDNA clones of rat OBP_L.

Figure 2 shows a Northern blot analysis of the distribution of OBP_L message and OMP message in various tissues.

Figure 3 shows a Southern blot analysis of genomic DNA from rat liver using an OBP_L probe.

Figure 4 shows the amino acid sequence alignments of OBP_L and ten homologous proteins.

Figure 5 shows the *in situ* hybridization of labeled cRNA for OBP_L as well as immunohistochemical staining with anti-OBP_L antiserum and autoradiography using tritiated odorants.

Figure 6 shows a Western blot analysis of rat and cow nasal homogenates with anti-OBP_L antiserum.

DETAILED DESCRIPTION OF THE INVENTION

It is a discovery of the present invention that odorant-binding protein in the rat is synthesized in the lateral nasal gland. The rat has between fifteen and twenty discrete nasal glands. The lateral nasal gland is one which does not exist in the cow, but does exist in other mammals, such as the human.

Before the work of the present invention, the site of synthesis of odorant-binding protein in the rat was unknown. Odorant binding protein itself, as well as odorant binding activity, had been found in rat mucus and tears, as well as in the olfactory epithelium. Much less activity was found in the respiratory epithelium. In the cow, odorant-binding protein had been found immunohistochemically in the glands of the lamina propria and along the surface of the epithelium; it was found in both olfactory and respiratory glands.

It has now been discovered that the presence of OBP_L in the epithelium and mucus of rats is due to secretion from its site of synthesis in the lateral nasal gland. It would have been extremely difficult, if not impossible, to isolate the mRNA encoding OBP from these epithelial tissues in rats. Although not wishing to be

bound by any particular theory, applicants believe that it is only because they have used tissue preparations containing the lateral nasal gland to make their cDNA library, that they were able to isolate the OBP cDNA.

It has been found that the OBP_L gene is a member of a larger family of proteins which have ligand carrier properties. Table 1, below, shows the physical properties of the ligand-binding carrier family. The family membership of OBP_L has been recognized by use of computer comparison of amino acid sequences. All of the proteins in this family have a similar subunit molecular weight. Many of them bind and serve as carriers for small lipophilic molecules, while the function of some is yet undetermined.

Insecticyanin binds the small lipophilic molecule biliverdin IX. Apolipoprotein D binds cholesterol and cholesteryl esters and has been proposed as a carrier for cholesterol. One of the best characterized of these proteins is retinol binding protein. It serves as a carrier for retinol, transferring retinol from plasma to the pigmented epithelium of the retina.

One of the consensus sequences recognized among this family of proteins also occurs near the N-terminus of a different family of ligand-binding proteins of which retinol-binding protein is a member. Other members of this family include cellular retinol binding protein I and II, cellular retinoic acid binding protein, peripheral nerve myelin P2 protein, heart and intestinal fatty acid binding protein, and murine adipocyte protein P422.

While the inventors do not wish to be bound by any particular theory, it is proposed that OBP_L functions in an analogous fashion to retinol binding protein. Lipophilic odorants could more readily traverse the hydrophilic environment of the nasal passages to reach the odorant receptors on the cilia of olfactory neurons if bound to OBP_L. OBP_L might also facilitate the transfer of odorants to the receptors which are bathed in hydrophilic mucus.

According to the method of the present invention, an odorant-binding protein gene is isolated by preparing cDNA from mRNA which has been isolated from lateral nasal gland tissue. Since the lateral nasal gland is the sole site of expression of the odorant-binding protein RNA, it is beneficial that the tissue preparation contain predominantly lateral nasal gland cells. The lateral nasal gland is one of the largest of the nasal glands. See Bojsen-Moller, Anat. Rec., Vol. 150, pp. 11-24, 1964 and Cuschiere et al., Histochemical Journal Vol. 6, pages 543-558, 1974. The lateral nasal gland (LNG) is located in the posterior area of the nose, just ventral and anterior to the olfactory epithelium on the lateral wall. Its main excretory duct is long and extends anteriorly, opening into the internal ostium at the tip of the nose. Contamination of the lateral nasal gland tissue preparation with other tissues will reduce the frequency of OBP_L clones in the cDNA library isolated.

Messenger RNA can be isolated from lateral nasal gland tissue according to methods well known in the art. Briefly, RNA can be purified by affinity chromatography on oligo(dT) cellulose to separate the poly(A)-containing mRNA from other RNAs. Methods for (1) isolation of mRNA from mammalian cells, (2) selection of poly(A) RNA and (3) cDNA synthesis using mRNA as a template, can be found in Maniatis, et al. Molecular Cloning: A laboratory Manual, Cold Spring Harbor Labs, 1982.

The cDNA which has been prepared can be inserted into appropriate DNA vectors to form recombinant DNA molecules. Many methods are well known in the art for accomplishing this, for example, adding complementary homopolymer tracts to double-stranded cDNA and to the vector DNA. The vector and double-stranded cDNA are then joined by hydrogen bonding between the complementary homopolymeric tails. In vitro enzymatic ligation is not necessary to establish these recombinant vectors in *E. coli*. Alternatively, synthetic linkers can be added to the termini of the double-stranded cDNA. After cleavage with the appropriate restriction enzyme, the cDNA molecules are inserted into plasmid DNA that has been cleaved with a compatible enzyme. Again, such methods are well known in the art. See Maniatis, *Supra*.

Suitable host cells can be transformed according to known methods in the art to achieve uptake by the cells and stable replication of the recombinant DNA molecules. The cells may be of prokaryotic or eukaryotic derivation. Choice of suitable host cells and vectors is well within the skill of the art. *E. coli* is a common choice.

In order to isolate the transformed cells which contain a molecular DNA clone of the odorant-binding protein, a detection means must be employed. Again, choice of a detection means is well within the skill of the art, although a few means will be specifically mentioned below. In one detection method, a nucleic acid probe is used which comprises a mixture of sequences, each of which codes for a portion of the amino acid sequence of the odorant-binding protein. That is to say that the probe contains all of the possible coding sequences for a sequence of amino acids of odorant-binding protein. This method was used by the present inventors to construct a mixed oligonucleotide probe (21 mer) containing thirty-two distinct molecular species. These probes were constructed based on the frequency of codon utilization. See Lathe et al, *Journal of Molecular Biology*, Vol. 183, p. 1, 1985.

In another method for detecting odorant-binding protein genes, a nucleic acid probe is utilized which comprises at least about twenty nucleotides from the DNA sequence determined for rat odorant-binding protein. Thus the information gained from the rat odorant-binding protein gene can be used to isolate OBP_L genes from other mammalian organisms containing lateral nasal glands. Similarly, when OBP_L genes are isolated from other mammals, those sequences can be used to construct probes to still other mammalian species. In still another method for detecting cloned genes, cross-reacting material produced by the transformed cells is tested. That is to say that the products of transformed cells can be screened with antiserum which contains antibody reactive with odorant-binding protein of the rat. In cases where one has inserted the OBP_L DNA in a vector such that a fusion protein will be synthesized, or where full or partial length OBP_L gene expression is expected, this immunological method can be used. Methods for immunological as well as hybridization testing of recombinant clones are well known in the art.

In another detection method which can be used when cDNA has been inserted into DNA vectors such that expression can be expected, the transformed cells can be screened for the production of a protein which specifically binds odorant molecules. For example, binding of tritiated 2-isobutyl-3-methoxypyrazine (IBMP) can be used as an assay to identify transformed cells which bind IBMP, presumably by expressing an odorant-binding protein. See, for example, Pevsner et al., *Proceeding of the National Academy of Sciences U.S.A.*, Vol. 82 pp. 3050-3054, 1985.

Odorant binding protein can be made using host cells which have been transformed with recombinant molecules. The recombinant molecules have odorant-binding protein genes positioned downstream from a transcriptional promoter and downstream from and in phase with a translational start site. Suitable vectors and methods for constructing such recombinant molecules are well known in the art. Once such recombinant molecules and transformed cells have been produced, they can be grown in a culture medium under conditions such that the protein encoded by the odorant-binding protein gene is expressed. For example, if the odorant-binding protein gene is under the control of an *E. coli* lac promoter and operator, expression of the odorant-binding protein gene can be induced using isopropyl-beta-D-thiogalactoside.

The protein may be expressed as either the full-length odorant-binding protein, a portion thereof, or as a fusion protein with a portion of another protein covalently attached. Depending on the particular construction of the recombinant molecule, the protein which is produced may be harvested either from the growth medium (if the protein is secreted), or from the transformed cells themselves. Suitable methods for harvesting and isolating proteins from transformed cells and growth media are well known in the art.

DNA molecules which are contemplated and embraced by the present invention include those which are free of introns, because they have been isolated via mRNA, as discussed above. These cDNA molecules will have sequences which correspond to that shown in Figure 1 for rat OBPL. While this protein shares about 30% identity with other members of the family of lipophilic molecule carrier proteins, it also shares that amount of identity with pyrazine-binding protein isolated from the cow, (also called odorant-binding protein.) Thus OBPL isolated from the rat is no more similar to the pyrazine-binding protein of cow than it is to rat alpha-2-microglobulin or to rat major urinary protein. Nonetheless, it is expected that the odorant-binding proteins similar to OBPL will be found in other mammalian species, especially those of mammals having a lateral nasal gland. These homologous proteins would be expected to have substantial homology with OBPL, at least greater than about 50%, and preferably greater than about 75%.

While the overall protein structures of the OBPL family members (as determined by X-ray crystallography) are remarkably similar, none of the amino acids in the consensus sequences is in contact with the bound ligand. Thus, although applicants do not wish to be bound by any particular theory regarding the mechanism of action of OBPL, it is quite possible that odorants are bound by OBPL merely on the basis of their overall size, conformation, and hydrophobicity. This is consistent with the extremely catholic binding abilities found for OBPL. For example, OBPL will bind 2-isobutyl-3-methoxypyrazine, methylidihydrojasmonate, as well as 3,7-dimethyloctan-1-ol. Because of this extremely broad range of odorant binding, OBPL can be used for many different purposes.

In one method of use, OBPL is used to concentrate odorants on a solid support. OBPL can be covalently attached to the solid support or can be noncovalently attached, for example merely admixed or in intimate contact with the solid support such that the combination of OBPL and solid support is stable. Solid supports include human skin, agarose beads, cellulose, or other inert solids. In one such proposed method, the solid support is a human nose. If OBPL is applied to the exterior portions of the nose only, odorants will be concentrated outside of the nose and their diffusion into the nose to the olfactory perception system will be greatly reduced. Since the sense of taste and appetite are known to be enhanced by perception of certain odorants, such a method could be used to reduce the appetite of obese humans. Conversely, if OBPL were applied to the interior portions of the nose, for example, those portions bathed in mucus, odorants would be concentrated to the mucus and this would enhance the amount of odorant perceived by the human. This can be useful in situations where appetite has been depressed either by illness or chemotherapy. Further, applicants have also found that when OBPL is mixed with odorants, the odorant stimulation of the enzyme odorant-sensitive adenylate cyclase is enhanced relative to the stimulation in the absence of OBPL. Thus OBPL would both concentrate odorants and enhance their perception.

In still another method of use, OBPL protein is applied to human skin before or concomitantly with a perfume or perfumed product. By trapping the odorant of the perfume at the skin, OBPL can decrease the amount of such odorants absorbed by the skin. This would be beneficial in situations where it is not desirable that the odorants be absorbed into the skin; also it would prolong the useful time period of the perfume on the body.

The term perfume is used herein to mean a mixture of organic compounds including, for example, alcohols, aldehydes, ketones, nitriles, ethers, lactones, acetals, esters, and frequently hydrocarbons which are admixed so that the combined odors of the individual components produce a pleasant or desired fragrance. Also included here are natural and synthetic "essential oils". Such perfumes usually contain: (a) the main note or the "bouquet" or foundation-stone of the composition; (b) modifiers which round-off and accompany the main note; (c) fixatives which include odorous substances which lend a particular note to the perfume throughout all stages of evaporation, and substances which retard evaporation; and (d) top-notes which are usually low-boiling, fresh-smelling materials.

In yet another method of using OBPL, the protein is covalently attached to a chromatographic column packing material, for example, cellulose or agarose beads. A crude mixture of compounds can be added to

such a column packing material, and odorants are preferentially retained upon the column. The odorants can then be isolated, by specific elution from the chromatographic column material. Differential elutions from the chromatographic column also provide a means of separating one odorant from another. Such column chromatographic methods are well known in the art as well as methods of elution and differential elution.

Because OBP_L does bind odorants, it will have the effect of reducing the rate of volatilization of odorants from solutions and perfumed products. Thus, the use of OBP_L can prolong the effective shelf life of volatile odorant components in solutions and perfumed products. OBP_L will be mixed with such a solution or product in an amount which is volatilization-reducing. Once again, such amounts can be readily determined according to routine tests for measuring volatilization in the art. Perfumed articles generally comprise aqueous and non-aqueous media, such as detergents, fabric softeners, cosmetic powders, creams and lotions, colognes, as well as other articles known in the art. Cosmetic lotions and creams include preparations for cleansing, softening, smoothing, and protecting the skin. Lotions often are liquids containing one or more insoluble substances. Both lotions and creams may be suspensions or emulsions.

Because of its role as a lipophilic molecule carrier in a hydrophilic environment, OBP_L can also be used as a solubilizing agent. For example, a lipophilic odorant can be bound to OBP_L and then the bound mixture can be further mixed with an aqueous medium to form an aqueous perfumed product or cologne. Thus, OBP_L would increase the miscibility of lipophilic odorants in aqueous or semi-aqueous environments.

In still another method for using OBP_L, crude extracts of natural tissues can be screened for the presence of potent odorants. For example, a natural source, such as a fruit or vegetable, can be homogenized and extracted to form a crude extract. The crude extract can then be diluted over a broad range, for example, ten-fold dilutions over a range of six orders of magnitude. Each dilution can then be tested for its ability to displace the binding of a known strong odorant, for example, galbazine (IBMP), from OBP_L. For example, the strong odorant would be pre-bound to the OBP_L and preferably labeled to facilitate measurement. Any extract which is able to displace the strong odorant at a high dilution indicates the presence in the natural tissue source of a potent odorant.

The following examples are given to more clearly and specifically present the foregoing general description. The invention is not limited by the scope of the examples, but only by the claims which are appended further below.

Example 1

This example describes the cloning of the OBP_L gene.

Purified rat OBP_L was obtained using DEAE-cellulose chromatography and reverse phase HPLC, using the binding of tritiated odorants as an assay. See Pevsner et al, *Proceedings of the National Academy of Sciences U.S.A.* Vol. 82, Page 3050, 1985, and Vol. 83, Page 49-42, 1986. Direct N-terminal amino acid sequencing of the HPLC purified protein yields the sequence H₂N-ala-his-glu-asn-leu-asn-ile-ser-pro-ser-glu-val-asn-gly-asp. Based on the frequency of codon utilization, we constructed a mixed oligonucleotide probe (21 mer) containing thirty-two distinct sequences. This probes corresponded to the seven N-terminal amino acids:

5' GC T -CA T -GA G -AA T -CTG -GA T -ATC-3'

A cDNA library in Lambda gt10 was prepared from male rat nasal epithelium containing lateral nasal gland, by the method of Gubler and Hoffman (*Gene*, Vol. 25, Page 263, 1983) and 80,000 recombinant clones were screened using the [³²P]-labeled oligonucleotide probes. Sixteen independent clones were isolated. The complete sequence was determined for three of these clones (see Figure 1). Hybridization-positive phage were subcloned in the plasmid vector Bluescript® (Stratagene San Diego, California) and sequenced by the Sanger random chain termination method (*Proceedings of the National Academy of Sciences U.S.A.* Vol. 74, Pages 5463, 1977.)

The three full length clones which have been obtained (i.e., that encode the full protein as it is isolated from rat tissue,) are clone 8F3 (801 base pairs), clone 850 (782 base pairs), and clone 800 (760 base pairs). All three clones have a 3' end at nucleotide number 801, but their 5' ends vary. Clone 8F3 begins at nucleotide number 1 in Figure 1. Clone 850 begins at nucleotide number 19, and clone 800 at nucleotide number 41. The coding sequence for the native protein as it is isolated from rat begins at nucleotide 62.

We detect a long open reading frame predicted to encode 172 amino acids. The predicted initiating methionine is followed by fourteen hydrophobic amino acids. The residues immediately following this presumed leader sequence correspond to the N-terminal protein sequence. Eight amino acids of the N-terminus determined by amino acid analysis were not encoded in the oligonucleotide probe. The fact that these eight amino acids in the cDNA correspond exactly to the sequence determined from amino acid analysis, confirms that the isolated cDNA indeed encodes OBP_L. The putative ATG start code at position 17 is surrounded by a consensus sequence typical of eukaryotic ribosomal start sites (Kozak, *Nucleic Acids Research* Vol. 12, Page 857, 1984,) and is indicated as boxed nucleotides in Figure 1. The amino acids which were determined from the protein sequencing are underlined in Figure 1.

Features of the determined sequence indicating that a full length cDNA clone has been isolated include: (1) a characteristic ribosome binding sequence at the putative initiating methionine; (2) a polyadenylation signal at

nucleotides 737 to 742; (3) a poly(A) sequence at the 3' end. Sequence analysis of a second clone reveals three, single-nucleotide substitutions compared to the first cloned sequence. All of these nucleotide changes occur in the 3' untranslated region. In Figure 1 these variations are indicated above the main line of sequence and marked with an asterisk. Bam HI, Taq I and Hinc II restriction sites are indicated by lines above the nucleotide sequence. The Hinc II site is present in clone 800 (the "variant" clone) but not in clones 850 or 8F3. The termination codon TAA is at position 553. The polyadenylation signal AATAAA is underlined. Both strands of clones 850 and 800 were completely sequenced. Clone 850 and 800 begin at positions 19 and 41 of clone 8F3, respectively.

Subclones of the larger cDNA clone from Lambda gt10 were recloned into Bluescript® phagemid's Eco RI site (obtained from Strategene Cloning Systems, San Diego, California). Bluescript® contains the T3 and T7 promoters in the coding region of the lac Z gene, thus allowing the production of fusion proteins. Between the T3 and T7 promoters is a multiple cloning site containing the Eco RI (and other) restriction sites.

The ten subclones which have been constructed in the Bluescript® phagemid contain the 5' and 3' Bam HI fragments of clone 850, the 5' and 3' Bam HI fragments of clone 800, the 5' and 3' Taq I fragments of clone 850, and the 5' and 3' Taq I fragments of clone 800. In addition, 5' and 3' Hinc II fragments of clone 800 have been inserted into Bluescript® phagemid. The latter fragment contains no translated DNA sequences of OBPL. All three of the large clones, 8F3, 850, and 800, contain all of the nucleotides which are necessary to code for the mature OBPL isolated from rat.

Example 2

This example shows Northern blot analysis indicating the tissue distribution of the mRNA for OBPL.

Total RNA was isolated from twelve tissues in the presence of guanidinium isothiocyanate as described in Thomas, Proceedings of the National Academy of Sciences U.S.A., Vol. 77, p. 5201, 1980. RNA (15 ug per lane) was treated with glyoxal and subjected to electrophoresis on a 1% agarose gel in 10 mM sodium phosphate buffer (pH 7.0). RNA was transferred to Nytran® (Schleicher & Schuell) and hybridized to [³²P]-labeled clone 850 (specific activity 6 x 10⁸ dpm/ug) by random primer labeling. The filter was hybridized for twelve hours at 42°C, washed twice for twenty minutes 0.3 M NaCl/30 mM sodium phosphate/0.5% SDS at 22°C, washed for twenty minutes in 75 mM NaCl/7.5 mM sodium phosphate/0.5% SDS at 42°C, air dried, and autoradiographed. Molecular sizes were determined from a one kilobase ladder (obtained from BRL, Bethesda, Maryland) subjected to electrophoresis in adjacent lanes.

The tissues which were tested for the presence of hybridizing RNA were olfactory epithelium, lateral olfactory epithelium including the lateral nasal glands, nasal respiratory epithelium, and olfactory bulb. Detectable signal was apparent only in nasal tissues, indicating that mRNA for OBPL is restricted to the nose. The abundance of OBPL message is comparable in respiratory and olfactory epithelium. However, the highest level of message are detected in the region of the lateral nasal gland (LNG), also known as Steno's gland. Microscopic localisation of OBPL mRNA by in situ hybridization reveals a highly selective localization of OBPL message to the LNG.

The OBPL mRNA observed in olfactory and respiratory epithelium might reflect contaminating LNG tissue. The size of the OBPL message is about one kilobase, which is 200 nucleotide longer than the size of the cloned cDNA. The significance of the slightly higher molecular weight of the message in the respiratory epithelium compared to the LNG and the olfactory epithelium is not clear, but may reflect a differential extent of polyadenylation.

Example 3

This example shows a Southern analysis of rat genomic DNA using an OBPL probe.

Genomic DNA was isolated from rat liver and digested for ten hours at 37°C with Hinc II (lanes 2 and 4 or Bam HI (lanes 1 and 3.) See Figure 3. Fifteen ug of DNA per lane were subjected to electrophoresis on a 1% agarose gel and transferred to Nytran®. Clone 800 was digested with Hinc II to generate a 5' fragment (corresponding to nucleotides 41 to 570 of clone 8F3). Genomic DNA blots were hybridized for twelve hours at 65°C to [³²P]-labeled clone 800 (in panel a) or the 5' cDNA fragment (in panel B). Filters were washed twice for fifteen minutes in 0.3 M NaCl/30 mM sodium phosphate/0.1% SDS at room temperature; then washed for ten minutes in 75 mM NaCl/7.5 mM sodium phosphate/0.1% SDS at 65°C, air dried, and autoradiographed. Molecular sizes were determined from lambda/Hind III markers (obtained from BRL) subjected to electrophoresis in adjacent lanes.

The results suggest a limited family size of about two to four members. Because of the differences between the two OBPL cDNA clones in the 3' untranslated region, it was thought that at least two genes would be in the OBPL family. These results show that microheterogeneity in the OBPL protein is not responsible for the ability of OBPL to bind odorants of widely varying chemical structures. Such a microheterogeneity among an apparently homogeneous OBPL was formally possible before these results were obtained.

Example 4

The sequence of OBPL obtained from the cDNA clones was compared by computer to other sequenced proteins. A similarity was revealed between OBPL and a group of proteins with similar properties. See Table 1, and Figure 4. See Pervaiz et al., Science, Vol. 228, p. 335, 1985; and FASEB Journal, Vol. 1, p. 209, 1987; and Sawyer, Nature, Vol. 327, p. 659, 1987.

Homologies within the family of proteins related to OBPL occur at several sites which are indicated by the shaded areas in Figure 4. Two consensus sequences are apparent, beginning at amino acid residues 12 and 89 of mature OBPL. The first is basic-hyd-X-gly-X-trp-aro-X-hyd where basic is arg or lys, hyd is a hydrophobic residue (ile, leu, met, phe, tyr), aro is aromatic (phe or tyr) and X is any residue. The second consensus sequence is aro-X-hyd-hyd-X-thr-asp-tyr-asp-X-aro-hyd-hyd. The hyd-X-gly-X-trp consensus sequence also occurs near the N-terminus of a different family of ligand-binding proteins of which retinol-binding protein is a member. Other family members include cellular retinol binding proteins I and II, cellular retinoic acid binding protein, peripheral nerve myelin P2 protein, heart and intestinal fatty acids binding protein, and murine adipocyte protein P422. We suggest that these two families are evolutionarily related.

The structures of retinol-binding protein, beta lactoglobulin, and insecticyanin have been determined by X-ray crystallography at high resolution. These three proteins share a remarkably similar structure containing an eight-stranded antiparallel beta-barrel motif that forms the core of the molecule. The first consensus sequence (containing gly-X-trp) occurs near the end of the first beta-barrel, while the second sequence (containing thr-asp-tyr) occurs near the sixth beta-sheet. None of the amino acids in the consensus sequences is in contact with the bound ligand.

Initial alignments were made using the DFASTP program of Lipman and Pearson, Science, Vol. 227, p. 1435, 1985, on a VAX/VMS computer; final alignment was manual. Abbreviations and sources of protein sequence data which are used in Figure 4 are: alpha 2M=alpha2 microglobulin (Unterman et al., Proceedings of the National Academy of Sciences U.S.A., Vol. 78, p. 3478, 1981); BBLG=Bovine beta-lactoglobulin, (Preaux et al., Hoppe-Seyler's Z. Physiol. Chem., Vol. 360, p. 1595, 1979); BG=protein BG of Bowman's glands, (Lee et al., supra); HC=protein of heterogeneous charge (Takagi, Biochemistry and Biophysics Research Communications, Vol. 98, p. 997, 1981, and Lopez et al., Biochemistry and Biophysics Research Communications, Vol. 103, p. 919, 1981); AGP=Alpha1-acid glycoprotein (Ricca et al., Journal of Biological Chemistry, Vol. 256, Page 11199, 1981); APOD=apolipoprotein D, (Drayna et al., Journal of Biological Chemistry, Vol. 261, p. 16535, 1986); Thi=Tobacco Hornworm Insecticyanin (Riley et al., Journal of Biological Chemistry, Vol. 259, p. 13159, 1984); RBP=Retinol Binding Protein (Colantuoni et al., Nucleic Acids Research, Vol. 22, p. 7769, 1988 and Pervaiz et al., FASEB Journal, Vol. 1, Page 209, 1987); Purp=Purpurin (Burman et al., Cell, Vol. 51, p. 135, 1987). The amino acid residues in Figure 4 are shaded where seven or more residues in a column are either identical, acidic, basic, aromatic, or hydrophobic.

Example 5

This example demonstrates the localization of OBPL mRNA to coronal sections of rat nasal epithelium.

Two RNA probes were produced by transcription from a rat OBPL cDNA template. The cDNA containing 760 nucleotides including all of mature OBPL as well as part of its predicted hydrophobic leader sequence and a 3' untranslated region was subcloned in the plasmid vector pGEM-2 (Promega Biotech, Madison, Wisconsin). Run-off complementary RNA (cRNA) transcripts were generated using the SP6 polymerase to form antisense probes (complementary to mRNA) or using the T7 polymerase to form sense probes (identical to mRNA). The cDNAs were made according to the protocol of Promega Biotech using the restriction enzymes in Hind III and Sph I to yield run-off transcripts of 860 (antisense) and 1300 (sense) nucleotides. Transcripts were labeled with [³⁵S]-CTP to a final specific activity of 1,250 Ci/mmol.

The *in situ* procedure was modified from the method of Uhl. In *Situ Hybridization in Brain*, Plenum, New York, 1986. Male rats (200 grams, Sprague Dawley) were perfused with 0.1% paraformaldehyde in phosphate-buffered saline (PBS) and whole nasal epithelium was embedded in Tissue-tek® (Miles Laboratories, Naperville, Illinois). The tissue was rapidly frozen and 10 um coronal sections were cut on a cryostat at -20°C. Sections were post-fixed for five minutes in 4% paraformaldehyde in PBS, rinsed twice in sterile PBS, incubated for ten minutes at 23°C in 0.25% acetic anhydride/0.1 M triethanolamide/0.9% NaCl and then dehydrated through a graded series of ethanol (70%, 80%, 95%, 100%), chloroform (100%), ethanol (100%, 95%).

Sections were then incubated with antisense or sense labeled cRNA probes (1 x 10⁶ cpm per section) at 38°C for 18 to 24 hours. After hybridization, sections were washed four times for fifteen minutes in 1X SSC (150 mM NaCl/15 mM sodium citrate) at 55°C. To confirm the hybridization specificity, antisense cRNA was coincubated with 1700-fold excess unlabeled cRNA. Additional sections were post-hybridized with RNase A (12.5 ug/ml 22°C, 10 minutes).

In situ hybridization reveals a striking density of the labeled cRNA for OBPL in the LNG. See Figure 5a. Hybridization of the antisense probe to tissue sections results in a high density of silver grains over the LNG. This labeling is specific for OBPL mRNA, since incubation with the sense probe produces virtually no silver grains, see Figure 5B. The specificity of labeling of OBPL mRNA is further substantiated by the elimination of silver grains with excess unlabeled probe. A similar pattern of localization is observed when silver grains are visualized with emulsion-coated covered slips.

Example 6

This example demonstrates localization of OBPL by immunohistochemistry.

OBPL was purified from adult rat olfactory epithelium using tritiated odorant binding as an assay. Antibodies were raised in rabbits by standard techniques. Cryostat tissue sections were prepared as described above except that rats were perfused in 1% glutaraldehyde. Sections were immunohistochemically stained using an avidin/biotin/peroxidase technique (Vector Laboratories) using diaminobenzidine and hydrogen peroxide as

substrates. Tissue sections were incubated with a 1:12,000 dilution of antiserum against rat OBPL or preimmune serum for 48 hours at 4° C.

Immunohistochemical staining with this antiserum reveals a selective localization of OBPL in the rat to the LNG (see Figure 5c). No staining is apparent with pre-immune serum. (See Figure 5d)

Example 7

The example demonstrates the specificity of an antibody raised against OBPL.

We have raised polyclonal antibodies to OBPL. Western blot analysis of rat nasal homogenates reveals that this antiserum recognizes a doublet with subunit molecular weight of about 20 kDa, corresponding to the molecular size of mature OBPL. See Figure 6. No OBPL immunoreactivity is apparent in bovine nasal homogenates, indicating a lack of immunological cross-reactivity between bovine OBPL and rat OBPL.

Example 8

This example demonstrates the localization of odorant binding activity by tritiated odorant autoradiography.

Twenty day old male rats were perfused with 0.4% paraformaldehyde. Tissues were embedded in 50% brain paste/50% Tissue-Tek®, rapidly frozen at -20° C., and 8 µm cryostat tissue sections were cut and desiccated on slides for twelve hours. Slides were preincubated for five minutes in 30 mM Tris-HCl (pH 7.4) then incubated for 90 minutes at room temperature in 100 nM tritiated IBMP, tritiated amylacetate, or tritiated isovaleric acid, or in 200 nM tritiated DMO. Nonspecific binding was determined by co-incubation of the radioactive odorant with 500 µM unlabeled IBMP, amylacetate or isovaleric acid or with 50 µM DMO. After incubation, slides were rinsed in 30 mM Tris-HCl (30 seconds), washed twice for one minute in water, dried, desiccated for twelve hours and then exposed to Ultrafilm® (Kodak) for three weeks at -20° C.

We have localized OBPL in rat nasal tissue by autoradiography utilizing tritiated odorants. See Figure 5e. Autoradiographic grains are displaced by an excess of unlabeled odorants, indicating that the binding is saturable. See Figure 5f. The bound tritiated odorants are confined to the LNG with a pattern essentially the same as that observed by OBPL immunohistochemistry and *in situ* hybridization. We have observed saturable autoradiographic patterns with the odorants IBMP, DMO, and amylacetate which all bind to purified bovine OBPL and rat OBPL. Tritiated isovaleric acid, which fails to bind bovine OBPL or rat OBPL, also fails to specifically label the nasal epithelium autoradiographically. This further confirms the specificity of the odorant autoradiography for OBPL.

Claims

1. A method of isolating an odorant-binding protein gene comprising:
preparing cDNA using an mRNA template isolated from lateral nasal gland tissue;
inserting the cDNA into DNA vectors to form recombinant DNA molecules;
transforming host cells with said recombinant DNA molecules to form transformed cells; and
detecting the presence of odorant-binding protein DNA in said transformed cells.
2. The method of claim 1 wherein said detecting of odorant-binding protein DNA is accomplished by hybridization of a nucleic acid probe to the DNA of said transformed cells, said probe containing sequences which, when translated, produce the amino acid sequence of an odorant-binding protein.
3. The method of claim 1 wherein detecting the odorant-binding protein DNA is accomplished by hybridization to a nucleic acid probe which comprises a sequence of at least about twenty nucleotides from the sequence of the rat odorant-binding protein gene.
4. The method of claim 1 wherein detecting said odorant-binding protein DNA is accomplished by testing said transformed cells for the production of protein which is immunoreactive with anti-odorant-binding protein antibody.
5. The method of claim 1 wherein detecting said odorant binding DNA is accomplished by testing said transformed cells for the production of a protein which specifically binds odorant molecules.
6. A method of producing an odorant-binding protein, comprising:
transforming host cells with a recombinant molecule comprising an odorant-binding protein gene which is positioned downstream from a transcriptional promoter and downstream from and in phase with a translational start site to form transformed cells;
growing said transformed cells in a culture medium under conditions such that said transformed cells express the protein encoded by said odorant-binding protein gene; and
harvesting the expressed protein encoded by said odorant-binding protein gene from said transformed cells and/or said growth medium.
7. A DNA molecule which is free of introns and which encodes a protein having odorant binding abilities and having a sequence substantially as that shown in Figure 1.
8. A method of concentrating odorants on a solid support comprising:
applying an OBPL to a solid support; and
contacting said solid support with odorant compounds to effect binding of the odorant to said solid support via OBPL.
9. The method of claim 8 wherein the OBPL is covalently attached to the solid support.

10. The method of claim 8 wherein the OBP_L is applied as a liquid solution to the solid support and the liquor of said solution is evaporated to provide a solid support with a residue of OBP_L dried on said solid support.

11. The method of claim 8 wherein the OBP_L is in a solid form and admixed with the solid support.

12. The method of claim 8 wherein the solid support is a human nose.

13. The method of claim 12 wherein the OBP_L is applied to the exterior portions of the nose only so that odorants are concentrated outside of the nose to reduce the amount of odorant perceived by the human.

14. The method of claim 12 wherein the OBP_L is applied to the interior portions of the nose so that the odorants are concentrated inside the nose to enhance the amount of odorant perceived by the human.

15. The method of claim 10 wherein the solid support is human skin and a perfume or perfumed product is also applied to the human skin.

16. The method of claim 9 wherein the solid support is chromatographic column packing material.

17. A method of reducing the rate of volatilization of odorants from solutions and perfumed products, comprising:

providing a solution or perfumed product containing a volatile odorant; and
mixing a volatilization-reducing amount of OBP_L with said solution or perfumed product.

18. A method of solubilizing lipophilic odorants in aqueous media comprising:

providing an aqueous medium;

binding a lipophilic odorant to OBP_L; and

mixing said OBP_L-bound lipophilic odorant with the aqueous medium to form an aqueous perfumed product.

19. A method of screening for potent odorants in crude extracts of natural tissues, comprising:

binding a strong odorant to OBP_L to form bound OBP_L and strong odorant;

making an extract of a natural tissue;

making serial dilutions of the extract; and

determining which dilutions of the extract are able to displace the strong odorant from OBP_L, an extract which can displace the strong odorant at high dilution indicating the presence of a potent odorant in the natural tissue.

FIG. 1

1 G AAT CCA GGC TCT AAC ATG GTG AAG TTT CTG CTG ATT GTT CTT CCA TTA GGT GTA TCC TGT GCA CAT CAT GAA 19
 MET Val Lys Phe Leu Leu Ile Val Leu Ala Leu Gly Val Ser Cys Ala His Glu
 74 AAT CTT GAT ATC AGT CCC TCA GAG GTT AAT GGG GAC TGG CGC ACC CTT TAC ATA GTT GCA GAT AAT GTG GAG AAG 44
 Asn Leu Asp Ile Ser Pro Ser Glu Val Asn Gly Asp Trp Arg Thr Leu Tyr Ile Val Ala Asp Asn Val Glu Lys
 Bam HI
 149 GTA GCA GAA GGT GGA TCC CTG AGA GCT TAC TTT CAG CAC ATG GAA TGT GGT GAT GAA TGC CAG GAA CTC AAA ATC 69
 Val Ala Glu Gly Ser Leu Arg Ala Tyr Phe Gln His Met Glu Cys Gly Asp Glu Cys Gln Glu Leu Lys Ile
 224 ATA TTC AAT CTC AAG TTG GAC AGT GAA TCT CAG ACA CAC ACT GTT GTG GGA CAA AAA CAT GAA GAT GGG CGG TAC 94
 Ile Phe Asn Val Lys Leu Asp Ser Glu Cys Gln Thr His Thr Val Val Gly Gln Lys His Glu Asp Gly Arg Tyr
 299 ACT ACT GAC TAC TCT GGT AGA AAT TAC TTC CAT GTT TTG AAG ACA GAT GAC ATT ATT TTC TTT CAC AAC GTT 119
 Thr Thr Asp Tyr Ser Gly Arg Asn Tyr Phe His Val Leu Lys Lys Thr Asp Asp Ile Ile Phe Phe His Asn Val
 Tag I
 374 AAT GTC GAT GAG ACT GGA AGG AGA CAA TGT GAT TTA GTT GCT GGG AAA AGA GAG GAC CTG AAC AAA GCA CAG AAG 144
 Asn Val Asp Glu Ser Gly Arg Arg Gln Cys Asp Leu Val Ala Gly Lys Arg Glu Asp Leu Asn Lys Ala Gln Lys
 449 CAG CAG CTT AGG AAG CTG GCT GAG GAG TAT AAT ATT CCA AAT CAG AAT ACC CAG CAC TTG CTG CCC ACA GAC ACT 169
 Gln Glu Leu Arg Lys Leu Ala Glu Glu Tyr Asn Ile Pro Asn Glu Asn Thr Gln His Leu Val Pro Thr Asp Thr
 A*
 524 TGT AAC CAA TAA AGA CTC CAT ATG GCT TCA CAA AGG ACA GCA AGG TCA GCA ATA TTT CCC ACA TCA CCT TTT CCA
 Cys Asn Gln stop
 599 TGA AAT CAG AAT CGT GAC AAT GAA GAT AAC TCA TCC TTT TCT TAT TTT TTC TCT TCA TCT TTC CTA TGA AGC CAG
 674 AAA ATC TGC TTC GTG GAT TTG TTT CCC ACC CTC CTA TCA TGG TAC TGA TTC TTC TGT TGA TAA AAT AAA TTT ATT 194
 G*
 749 TTT CAT GCA CAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AAA AA 801

*Clone 800

FIG.2A

liver
lung
kidney
heart
intestine
↓olfactory
↓lateral olf
↓respiratory
olf bulb
cerebrum
salivary glands
harderian gland

FIG.2B

FIG. 3

FIG. 3

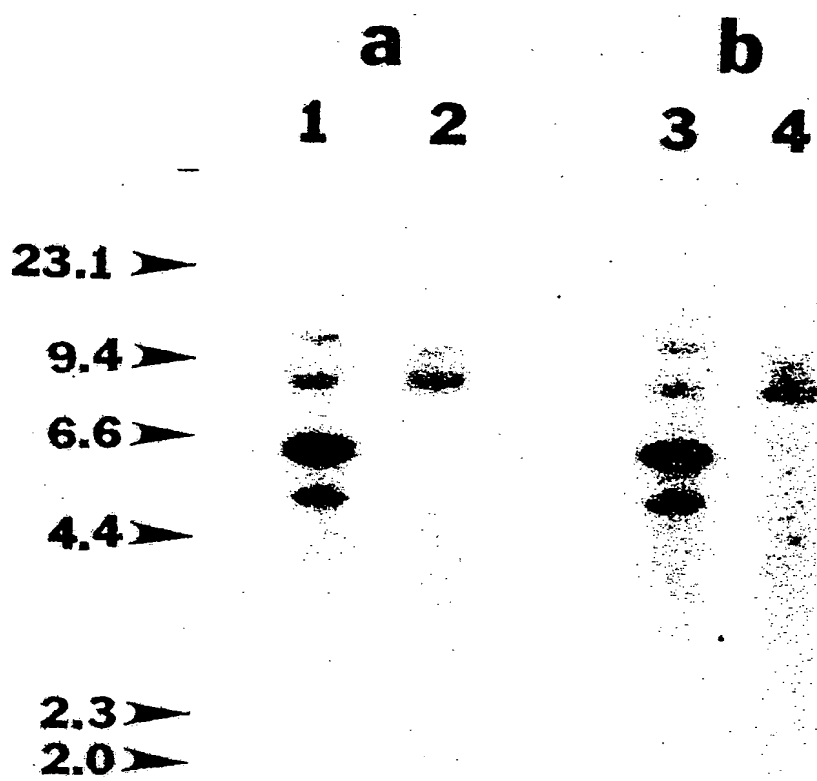


FIG. 4

OBP	MVKFLLIYV LALGVSCAHHENLDISPS	26
A2G	LLLLCLGLTLV-CGHAEASSTRGNLDVA	
BBLG	LIVTOTMKGLDIO	
BG	QC-OADLPPVMKG-LEEN	
HC	GPVPTPPDNIOVOENFNIS	
APOD	MVMLLLLLLSALAGLFGAAEGCAFHLGKCPNPPVOENFDVN	
THI	GDIFYPGYCPDVKPVNDFDLS	
RBP	MKWWWALLLLAAWAAAERDC-RVS-SFRVKENFDKA	
PURP	MKYAOYVFLASIFSAVEYSLAOTC-AV-DSFSVKDNFDPK	
AGP	MALHMLVVLSSLPLLEAONPEPANITLGIPITNETLK	
OBP	EVNGDWRTLYIVAD-NVEK--VA-EGGSLRAY-FOHMECG	61
A2G	KLNGDWFSIVVASN-KREK--IE-ENGSMRVF-MOHIDVL	
BBLG	KVAGTWYSLAMAAS-DISL--LDAOSAPLRVY-VEELKPT	
BG	KVTGVWYGI AAASN-CKOF--LOMKSDNMPAP-VNIYSLN	
HC	RIYGKWNLAIGSTCP-LK--IM-DRMTVSTL-VLGEGAT	
APOD	KYLGRWYEIEKIPPTTFENGRCIOA-NYSL--MENGKIKVL	
THI	AFAGAWHEIAKLPL-ENEN---O-GKCTIAEY-KYDGKKA	
RBP	RFSGTWYAMA-KKD-PEGL--FL-ODNIVAEFVSDETGOM	
PURP	RYAGKWYALA-KKD-PEGL--FL-ODNISA EYTVEEDGTM	
AGP	WLSDKWFYMGAAFRDPVFK--QA-VOTIOTEY-FYLTTP--	
OBP	-DECOELKII FNVKLD-SECOTHTVVGO-K-HEDGRY-T-	95
A2G	-ENSLGFK--FRIKEN-GECELYLVAY-KTPEDGEY-F-	
BBLG	PEGDLEIL--LOKWEN-GECAOKKIIAE-KTKIPAVF-K-	
BG	-NGHMKSS--TSFOTE-KGCCQMD-VEM-TTVEKGHYKW-	
HC	-EAEISMT--STRWRK-GVCEETS-GAYEKTDTDGKFLYH	
APOD	-NOELRAD--GTVNOIEGEATPVNLTEPAK--LEVKF---	
THI	-SVYNSFV-SNGVKEY-MEG-DLEIAPDAKYTKOGKY-V-	
RBP	-SATAKGR--VRLLNWDVYCADM-VGTFTDTEDEPAKF-KM	
PURP	-TASSKGR--VKLFGFWVICADMAAOYTVPDPTTPAKMYM	
AGP	-NLINDTIELREFOTTDDOCVYNFTHLG-VORENGTL-S-	
OBP	-TDYS-GR-NY-----FHV LKKT-DDIIF-FH-NVNV-DES	124
A2G	-VEYD-GG-NT-----FTILKTDYDRYVM-FH-LINF-KNG	
BBLG	-IDAL-NE-NK-----VLVLDTDYKKYLL-FC-MENS-AEP	
BG	-KMOO-GD-SE-----TII VATDYDAFLH-EF-TKIO-M-G	
HC	KSKWN-ITMES-----Y-VVHTNYDEYAJ-FL-TKKF-SRH	
APOD	-SWFM-PS-AP-----YWILATDYENYAL-VY-SCTC-IIO	
THI	-MTFKFGORVVNL-VPWVLATDYKNYA INYNCDYHPDKKA	
RBP	KYWGVASFLQKGNDDHWIVDTDYDTYAVQY-SCRLN-LD	
PURP	TYOGLASYLSSGGDNVWVIDTDYDNYAITY-ACRSLK-ED	
AGP	-KCA--GAVKI-----FAHLIVLKKHGTf-ML-AFNL-TDE	
OBP	G-RROCDL-VAGKREDLNKAOKOELRKLAEEYNIPNENTO	162
A2G	E-TFOLMV-LYGRTKDLSSDIKEKFAKLCEAHGITRDNII	
BBLG	EOSLACQC-L-VRTPEVDDEALEKFDKALKALPMHIRLSF	
BG	A-EVCVTVKLFGRKDTLPEDKIKHFEDHIEKVGLKKEOYI	
HC	H-GPTITAKLYGRAPOLRETLLODFRVVAOGVGIPEDSIF	
APOD	-LFHVDFAWILARNPNLPPEVDS LKNILTSNNIDVKKMT	
THI	H-SIHAWI-L-SKSKVLEGNTKEVVDNVLKTFSHLIDASK	
RBP	GTCADSYSFVFSRDPNGLPPEAOKIVROROEELCLAROYR	
PURP	GSCDDGYSLIFS RNPRGLPPAIORIVROKOE EICMSGOFO	
AGP	---NRGLS-FYAKKPDLSPELRKIFOOAVKDVGMDESEIV	

FIG. 4 (cont.)

OBP	HLVPTDT-CNO 172
A2G	DLTKTDR-CLOARG
BBLG	NPTOLEEOCHI
BG	RFHTKAT-CVPK
HC	TMADRGE-CVPGEOEPEPILIPR
APOD	-VTDOWN-CPKLS
THI	FISNDFSEAACOYSTTYS LTGPDRH
RBP	LIVHNGY-CDGRSERNLL
PURP	PVLCSGA-C
AGP	FVDWTKDKCSEOOKOOLELEKETKKETKKDP

FIG. 5

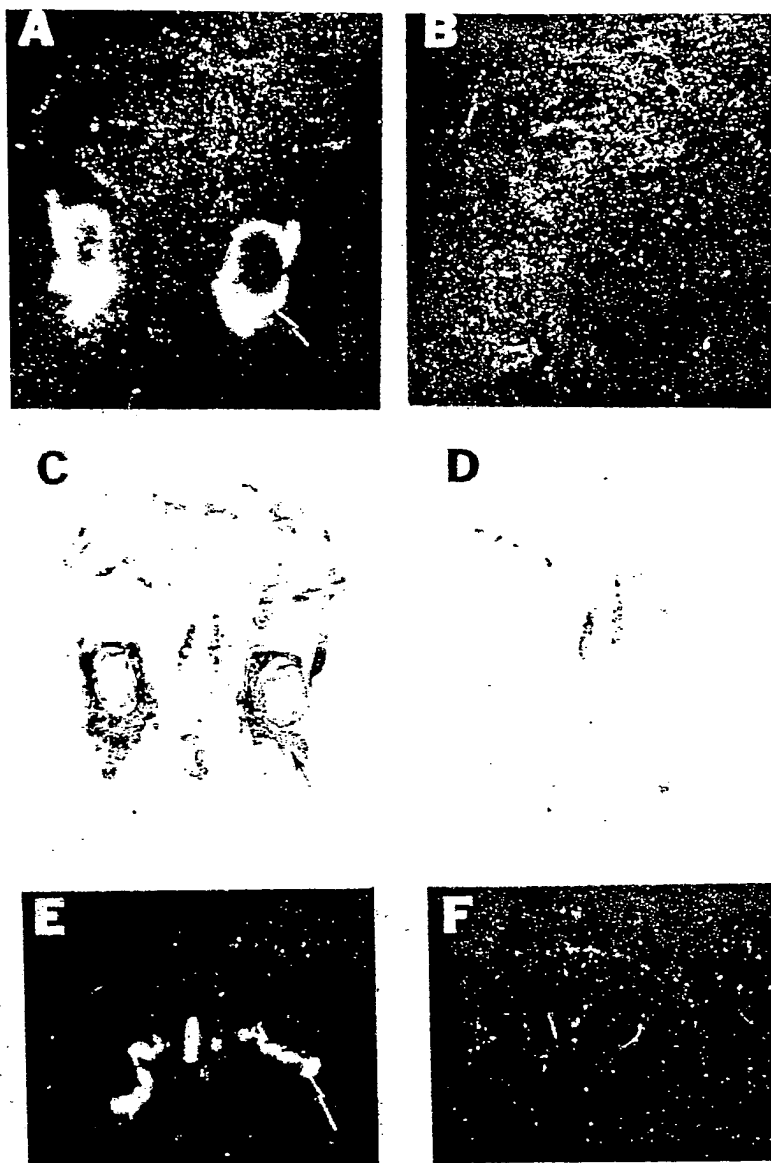


FIG. 6

